Original Article

Evaluation of Therapeutic Potentials of α-Lipoic Acid Versus Stem Cells-Derived Microvesicles Against Experimentally–Induced Gastric Ulcer in Adult Male Albino Rats (Light and Electron microscopic study)

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ABSTRACT

Background: Alpha-Lipoic acid (ALA), is an antioxidant endogenous substance. It is available in small amounts in food. Mesenchymal stem cells (MSCs) derived microvesicles (MSC-MVs) have numerous paracrine protective effects against tissue injury that are mediated by inflammatory, apoptotic and oxidative stress.

Aim of the Work: Our study was done to assess the protective effects of Alpha Lipoic Acid versus MSC-MVs on indomethacin-induced gastric ulcer in rats.

Material and Methods: Forty adult male rats were divided into: Group I (10 rats) represented control animals, and Group II (30 rats) involved ulcerated rats. The ulcerated group received a single gavage of indomethacin (30mg/kg B.Wt.) and was equally subdivided into: Subgroup II-a (ulcerated subgroup), subgroup II-b ulcerated rats that received a single intravenous dose of MSCs-MVs (0.5 mg/mL) and subgroup II-c ulcerated rats that received ALA (100 mg/kg) orally for 3 days before ulcer induction. Antioxidant enzymes and ulcer index were assessed. Gastric tissue was examined by light and transmission electron microscopes.

Results: Indomethacin caused marked damage of the gastric mucosa and ulcer index was significantly higher as compared to the control group. There was loss of the normal histological architecture and inflammatory cells infiltration. Ultrastructure sections revealed increased mucin granules in surface mucous secreting cells. Parietal cells showed dilatation of intracellular canaliculi. In ALA and in MSCs-MVs treated rat groups, there was a significant increase in antioxidant enzyme activities. The structure of the mucosa was significantly improved with a significant decrease in gastric ulcer area. Erosions, necrosis and inflammatory cell infiltration were significantly decreased. No major damage of endothelial cells was observed. The parietal cells count and the VEGF -positive cells were significantly decreased as compared with the control group.

Conclusion: MSCs-MVs have significant superior therapeutic effects on gastric mucosa against indomethacin-induced gastric ulcer than the protective effects of α-Lipoic acid (ALA).

INTRODUCTION

Gastritis is inflammation of the stomach mucosal lining. It can range from mild, asymptomatic to severe ulcerative form, which if untreated could lead to perforation¹. Peptic ulcer diseases (PUDs) are painful sores or ulcers that affected the lining mucosa of the stomach or the first part of the duodenum². Major complications of PUDs are bleeding, penetration, perforation or obstruction³⁴. Numerous causes that induce development of PUDs include: H. pylori infection, NSAIDs intake, smoking, alcoholism, Zollinger-Ellison syndrome, family history and others³⁵.

“Indomethacin induced damage” is considered as standard model of gastric ulcer in rats in order to study pharmacological and pathophysiological effects¹. Prokaryotic and eukaryotic cells are the main site of Alpha lipoic acid (ALA) as it acts as an effector in the mitochondrial enzymes pathway. Also, ALA is present in all types of foods especially in spinach, yeast extract, liver, and kidney⁶. Alpha-Lipoic acid (ALA) has beneficial effect as it is considered a strong antioxidant so, it can be used to protect and to treat many diseases⁷⁸.

On the other hands, (MSCs) derived micro vesicles (MSCs-MVs) mediate various biological effects and might be the main paracrine mechanism of communications between stem cells and injured cells. Yang et al.,⁹ evaluated the effects of MSCs-extracellular vesicles on experimental colitis. The therapeutic effects of MSCs-extracellular vesicles were mediated by the free radical scavenging...
activity, down regulation of the proinflammatory cytokine levels, inhibition of NFκBp65 signal transduction pathway and inhibition of the apoptosis[9].

The present study was conducted to compare and evaluate the antiulcerogenic and antioxidant effects of Alpha lipoic acid versus MSC-MVs on Indomethacin-induced gastric lesions in rats.

METHODS

Animal grouping

Male rats (40 animals, 150-250 g B. Wt) were obtained from an inbred colony in the animal house of Faculty of Veterinary Medicine, Benha University, Animals were kept at temperature (22 ± 2°C) and humidity (60%), with light: dark cycle alternates every 12 hours and free access to food and water. They received a balanced diet. All animal procedure was done accordance to the proper recommendations in the use and care of experimental animals. Animals were divided into two groups, Group I (Control group) involved 10 rats, Group II (ulcerated group) involved 30 rats that was equally subdivided into: Subgroup II-a (ulcerated subgroup), Subgroup II-b (ulcerated rats received MSCs-MVs). Animals received a single dose of MSCs-microvesicles (MSCs-MVs) at concentration of 0.5 mg/mL injected intravenously via the tail vein and Subgroup II-c (ulcerated rats received alpha lipoic acid).

Chemicals

1. Indomethacin (Sigma-Aldrich, MERCK, Cat. No. I7378-10G) was dissolved in distilled water. Two mL was given by intragastric gavage (30 mg/kg B.Wt.) to induce acute gastric ulcer[10].

2. Alpha lipoic acid (ALA) (Sigma Aldrich, MERCK Cat No. Q4951-10G) was dissolved in tween solution 0.2% (0.5 mL) before administration to animals ( tween is a non-ionic detergent used as an emulsifying agent in biochemical applications -Cat No.9005-64-5). Alpha- lipoic acid was delivered by oral gavage once daily (100 mg/Kg) for three consecutive days before induction of gastric ulcer by Indomethacin[10].

Isolation of MSC-derived Microvesicles

Microvesicles were separated from the supernatants of 1st, 2nd and 3rd passages of MSCs. Centrifugation at 2000 xg for 20 minutes was conducted to remove the debris. Centrifugation at 100,000 xg for 1 hour at 4oC was done for the cell-free supernatant by using ultracentrifuge of Beckman Coulter Optima L 90 K (AQ11). The pellet was washed by serum-free medium and by HEPES 25 mM (Sigma Aldrich, USA), then an ultracentrifugation was conducted for the 2nd pellet under the same conditions[12].

Identification and detection of MSC-derived Microvesicles

For detection of homing of MSCs-MVs into gastric tissue in rats, vesicles were labeled with Fluorescent PKH26 Red Cell Linker Kit (Sigma-Aldrich, Egypt) and injected into the tail vein of the Ulcerated group. Fluorescence microscope was used to examine the gastric tissue to visualize homing of PKH26 stained vesicles[13,14].

Macroscopic and Ulcer index study

All animals were fasted for 12-14 hours and allowed for drinking water only during the last day of experiment. Next day, scarification of all animals were done by cervical dislocation.

The stomach tissues were ligated around the cardiac and pyloric sphincters and filled with 3 mL distilled water. Gastric juice was collected in sterile tubes. Stomachs were incised longitudinally and washed with saline. Lesion in the glandular portion was examined under 10x magnifying lens to evaluate the ulcer formation, number of ulcers per stomach and the severity of ulcers. Mean ulcer score of every animal was stated as ulcer index. Curative percentage and ulcer index were calculated by using formulas previously described[15,16].

Biochemical Studies

Measurement of gastric enzymes

Gastric levels of glutathione (GSH), superoxide dismutase (SOD) and catalase (CAT) enzyme activities were assayed. Half gram of gastric tissue was ground in liquid nitrogen and homogenized in the appropriate buffer on ice for 15 minutes. Homogenates were used for assessment of enzyme activities according to the kit’s recommendations[17,18].

Glutathione (GSH) assay. Glutathione was determined by colorimetric detection kit (ThermoFisher Scientific, Cat. No. EIAGSHC) according to manufacturer’s recommendations. Results of the GSH were expressed as nmol/mg tissue[19].

Superoxide dismutase (SOD) and Catalase (CAT) assay

Commercial kits were used to assess SOD and CAT in stomach tissue according to manufacturer’s recommendations (Thermo-fisher Scientific, Cat No. EIASODC for SOD and Cat. No. EIACATC for CAT). Results were presented as mMol/min/mg tissue[20].

Histological study

Light microscopic study

Upper part of the fundus of stomach tissues was fixed in 10% formalin for preparation of Paraffin blocks. Serial
sections of 5 µm in thickness were subjected to HandE staining for histological details.

**Immunohistochemical study**

Immunohistochemical study for vascular endothelial growth factor (VEGF) (index for angiogenesis) was done by VEGF primary antibodies (monoclonal antibody from Novus Biologicals, USA, Cat. No. NB100-664SS). Citrate buffer and Mayer’s hematoxylin were purchased from Thermo-Scientific (Fremont, California USA). For antigen retrieval boiling of citrate buffer (10 mol/L) for 10 min was conducted. Sections were left for 20 min to cool down at room temperature. Primary antibodies were added to sections and incubation for 1 h was done. Secondary antibody was added (Biotin conjugated, Novus biologicals, USA, Cat No. MKB-2225-NB). Reaction was visualized using DAB; 3,3’-diamino-benzidine peroxidase substrate kit (Vector Labs, USA, Cat No. SK-4100). Mayer’s hematoxylin was used for counterstaining (Cat. No. TA-060-MH). Finally, Diaminobenzidine (DAB) chromogen solution was added and counter staining was done with Mayer’s hematoxylin. For the negative control section, the primary antibodies were excluded. Positive reaction for VEGF appeared in the form of brown cytoplasmic staining.

The protocol was previously described by University of Rochester Medical Center. A positive control was taken from a section of human placenta.

**Transmission electron microscopic study (TEM)**

Fixation in 2.5% phosphate-buffered glutaraldehyde (pH 7.4) was conducted for 1 mm of the upper part of the fundus of the stomach specimens. Stomach tissues were post fixed using 1% Osmium-Tetroxide at 4°C. The fixed gastric tissue samples were processed till embedding in Epon. Ultrathin sections (50 – 60 nm) were prepared using ultramicrotome. Sections were subsequently mounted on copper grids and stained with lead citrate and uranyl acetate. Ultrathin sections were assessed and photographed by using a JEM-1200EXII, TEM in Faculty of Science, Ain-Shams University.

**Morphometric study**

Image - Pro Plus program (version 6.0; Media Cybernetics Inc., Bethesda, Maryland, USA,) was used for morphometric study at the Pathology department, Faculty of Medicine, Cairo University for measuring: First; the mean number of cells lining fundic glands (surface cell, mucous neck cell, chief cell and parietal cell) from all experimental groups stained by hematoxylin and eosin-staining, 10 cells per section were observed (per high power field ) at X 40 objective from each animal of all groups and Second; the mean percentage of VEGF immunoexpression. Statistic–cal analyses were carried out using IBM SPSS statistics software for Windows (version 19; IBM Corp., Armonk, New York, USA).

**Statistical Analysis**

Results were exhibited as mean ± SE. Student paired t-test, one-way analysis of variance (ANOVA) and post-hoc tests were used for multiple sample groups comparisons. Statistical significance was considered at $p<0.05$. All tests were conducted by statistical package for social science (SPSS, version 19).

**RESULTS**

**Ulcet index result**

The ulcer index in group Ia was very high, 42.50±0.83 which confirm ulcer formation. The ulcer index for rats treated with MSCs-MVs was 17.53±0.86, a value significantly lower than that of group Ia and group Ic ($p<0.05$). However, rats taken α-Lipoic acid showed an ulcer index of 25.58±0.57 which was significantly lower than that of group Ia ($p<0.05$). (Table 1).

**Biochemical result**

**Effect on the antioxidant markers**

The antioxidant markers, GSH, SOD and CAT were significantly decreased in the indomethacin group when compared with the control group. Moreover, treatment with either lipoic acid or MSCs-MVs improved the antioxidant markers towards the normal group values. Alpha-Lipoic acid showed higher therapeutic effects as compared to MSCs-MVs (Table 2).

**Histological result**

**MSCs-MVs characterization and homing**

Transmission electron microscope revealed that the MSCs-MVs were rounded in shape (diameter less than 100 nm) (Figure 1A). Microvesicles labelled with PKH26 fluorescent dye were identified in vitro by a fluorescent microscope as strong red fluorescence (Figure 1B).

**Light microscope findings**

Stained sections by HandE of the control group showed normal architecture of the gastric mucosa. The gastric glands appeared narrow, numerous, straight, and perpendicular to the surface epithelium and filling the whole thickness of the mucosa. Smooth muscle fibers and blood vessels were present in the lamina propria. The gland has mucous cells (columnar with pale cytoplasm), parietal cells (central rounded nuclei) and chief cells in the basal region (columnar with basal oval nuclei, basal basophilic cytoplasm, and pale apical part) (Figure 2 AandB). The resulting mucosal lesions in subgroup Ia showed superficial damage with distorted shape of the gastric glands. The cells lining the damaged glandular area appeared shrunken with deeply acidophilic cytoplasm (Figure 2C). Basal part of the gland showed loss of architecture with inflammatory cells in the lamina propria with dilated and congested capillaries (Figure 2D).

Stained sections by HandE in subgroup Ib showed deep eosinophilic cytoplasmic epithelial cells with vacuolization of some cells, also increase in mucus secreting cells with inflammatory cells in lamina propria and widened gastric pits (Figure 2E).
In subgroup IIc sections showed that the gastric mucosa was more or less similar to the control group with inflammatory cells infiltration and congested blood vessels in the lamina propria (Figure 2F).

**Immunohistochemical result**

Stained sections by VEGF of the control group showed moderate positive reaction for VEGF in the cells of the fundic glands (Figure 3A), while in subgroup IIa showed negative reaction for VEGF compared to the control group (Figure 3B). Both subgroup IIb and subgroup IIc showed strong positive reaction for VEGF (Figure 3C and D).

**Electron microscope result**

Examination of the gastric mucosa of control group revealed normal ultrastructure of different cells covering gastric mucosa. The chief cells showed rounded euchromatic nuclei with smooth nuclear membrane and their cytoplasm showed mitochondria and apical pale zymogenic secretory granules (Figure 4A). Also, the gastric mucosa of the control group showed parietal cells that had numerous mitochondria, euchromatic nuclei with narrow intracellular canaliculi (Figure 4B) and the surface mucous-secreting cells showed rounded, multiple and electron-lucent mucous vacuoles in the cytoplasm with euchromatic nuclei. (Figure 4C).

The resulting electron microscopic examination of the mucosal lesions in subgroup IIa showed severe damage of the chief cells with small indented nuclei, few secretory granules and dilated cisternae of rough endoplasmic reticulum (Figure 4D). Some parietal cells had marked dilation of the intracellular canaliculi. Others showed electron-dense cytoplasm containing mitochondria (Figure 4E). Also showed surface mucous cells containing apical electron-lucent mucoid granules of different sizes with irregular nuclei (Figure 4F).

Examination of the gastric mucosa of subgroup IIb showed nearly the same ultrastructural features as the control group in which chief cells showed euchromatic nuclei, zymogenic granules and rough endoplasmic reticulum (Figure 5A). The parietal cells were nearly similar to control, having euchromatic nuclei, intact intracellular canaliculi with large number of mitochondria (Figure 5B). While the mucous cells in subgroup IIc showed different shapes of granules. The chief cells restored the normal shape of its nucleus and zymogenic granules (Figure 5C). Presence of a typical tubulovesicular appearance together with intracellular canaliculi was characterized in the cytoplasm of the parietal cells (Figure 5D).

**Morphometric result**

The mean count of different cells lining the fundic glands in control group were: mucous neck cells 8.46±0.48; parietal cells 11.16±2.24 and chief cells 36.84±2.04. There was non-significant difference (p value > 0.05) in the count of different cells lining the gastric glands after MSCs-MV administration in comparison to that of the control group as shown in (Table 3). In ulcerated group, most cells lining the fundic glands (mucous neck cells, parietal cells and chief cells) showed high significant decrease (p value 0.01) in count in comparison to that of the control. Treatment with both Alpha lipoic acid and MSCs-MV significantly restore the count of all cells that lining the gastric glands when compared with indomethacin group. In addition, there was non-significant decrease (p value > 0.05) in diameter of all cells lining the gastric glands after treatment with both Alpha lipoic acid and MSCs-MVs when compared with control group as shown in (Table 3).

The mean number of VEGF -positive cells showed a significant reduction in subgroups IIa as compared to the control group, whereas the other treated groups showed a significant elevation as compared to control group and indomethacin treated group as shown in (Table 3).
Fig. 2: (A) A photomicrograph of a section of the upper part of the fundus of the stomach from control group showing normal architecture of the gastric mucosa with gastric pits (P), lamina propria containing gastric glands (L) which appeared narrow, numerous tightly packed, straight, and perpendicular to the surface epithelium and occupying the whole thickness of the lamina propria. The lamina propria contained blood vessels and smooth muscle fibers (arrow), HandE X 200.

Fig. 2: (B) A section of the upper part of the fundus of the stomach from control group showing surface columnar mucous cells with pale cytoplasm (two arrow) and parietal cells (one arrows) with central rounded nuclei and eosinophilic cytoplasm, HandE X 400.

Fig. 2: (C) A section of the upper part of the fundus of the stomach from the indomethacin subgroup IIa showing epithelial erosion (arrow) with increased mucous cells with distorted nuclei and lightly stained cytoplasm (M). HandE X 400.

Fig. 2: (D) A section of the upper part of the fundus of the stomach from the indomethacin subgroup IIa showing loss of architecture of superficial epithelium and gastric pits (arrow). Mononuclear cellular infiltration in the lamina propria (L) with congested blood capillaries (V). Notice the damaged glandular area with loss of cellular arrangement (*). HandE X 400.

Fig. 2: (E) A section of the upper part of the fundus of the stomach from the MSCs-MVs subgroup showing almost normal gastric mucosa with wide gastric pits (P) and increase mucous cells (arrow). HandE X 400.

Fig. 2: (F) A section of the upper part of the fundus of the stomach in rats treated by alpha-lipoic acid subgroup showing almost normal gastric
macosa with normal gastric gland (arrows) HandE X 400.

**Fig. 3:** (A) A photomicrograph of a section of the upper part of the fundus of the stomach showing: (A) A moderate positive reaction for VEGF in the cells of the fundic glands of control group. VEGF immunostaining X 400.

![Photoelectron micrograph of a section of the glandular of the upper part of the fundic mucosa of a control rat showing pyramidal chief cell with basal rounded nucleus (N). zymogenic granules in the apical part of the cell (arrow). (E. M X 5000).](image1)

**Fig. 3:** (B) A negative reaction for VEGF in the cells of the upper part of the fundic glands of indomethacin subgroup IIa. (VEGF immunostaining X 400.

![A section of the upper part of the fundic mucosa of stomach of a control rat showing a parietal cell with a euchromatic nucleus (N), numerous mitochondria (M), and narrow intracellular canaliculi (I).](image2)

**Fig. 3:** (C) A strong positive reaction for VEGF (arrow) in the cells of the upper part of the fundic glands of MSCs-MVs subgroup. VEGF immunostaining X 400.

**Fig. 3:** (D) A strong positive reaction for VEGF (arrow) in the cells of the upper part of the fundic glands of alpha-lipoic acid subgroup. VEGF immunostaining X 400.

![A photomicrograph of a section of the upper part of the fundic glands of alpha-lipoic acid subgroup.](image3)

**Fig. 4:** (A) A photoelectron micrograph of a section of the glandular of the upper part of the fundic mucosa of a control rat showing pyramidal chief cell with basal rounded nucleus (N). zymogenic granules in the apical part of the cell (arrow). (E. M X 5000).

![A section of the upper part of the fundic mucosa of stomach of a control rat showing a parietal cell with a euchromatic nucleus (N), numerous mitochondria (M), and narrow intracellular canaliculi (I).](image4)
Microscopic magnification: 5000.

Fig. 4: (C) A section of upper part of the fundic mucosa of control rat showing surface mucous cell lining of gastric fundic gland with basal nucleus (N) and apical electron lucent secretory granules (S). E.M X 6000.

Fig. 4: (D) A photoelectron micrograph of a section of upper part of the fundic mucosal epithelial cells of stomach of indomethacin subgroup II showing chief cell with irregular heterochromatic nucleus (N), multiple degenerated zymogenic secretory granules (G) and vacuolation (V). Notice, degenerated rough endoplasmic reticulum (rER) in between the vacuoles. E.M X 7000.

Fig. 4: (E) A photoelectron micrograph of a section of upper part of the fundic mucosal epithelial cells of stomach of indomethacin subgroup II showing parietal cell with small irregular pyknotic nucleus (N), cytoplasmic vacuoles (V) with dilatation intracellular canaliculi (I).

Fig. 5: (A) A photoelectron micrograph of a section of the glandular of the upper part of the fundic mucosa of MSCs-MVs subgroup showing normal chief cell with intact nucleus (N), and apical zymogenic granules (G). (E.M X 7000).

Fig. 5: (B) A photoelectron micrograph of a section of the glandular of the upper part of the fundic mucosa of MSCs-MVs subgroup showing normal parietal cell showing nearly similar to control rounded euchromatic basal nucleus (N), numerous mitochondria (M) and extensive tubulovesicular...
**α-LIPOIC ACID VERSUS STEM CELLS MICROVESICLES IN GASTRIC ULCER**

**Discussion**

The pathogenesis of Indomethacin-induced gastric ulceration has been attributed to its inhibitory effects on prostaglandin synthesis as well as free radical formation [23]. Indomethacin as one of NSAIDs is frequently used as analgesic. Due to its inherent common side effects and the high cost of gastric ulcer pharmaceuticals, several studies evaluated the effect of natural products of plant origin which proved to be affordable, efficacious and non-toxic. The present study was conducted to evaluate the protective effects of Alpha lipoic acid versus MSC-MVs on indomethacin-induced gastric ulcer in rats.

The histological observations in the ulcerated subgroup IIa revealed areas of loss of superficial epithelium and ulceration. Moreover, there was destruction of the mucosal epithelial cells that showed pyknotic nuclei and highly eosinophilic cytoplasm that extended along the length of fundic glands from the neck to the base. These results agreed with previous studies [24, 25]. Moreover, the antioxidant markers; GSH, SOD and CAT were significantly decreased. These results explain that indomethacin-induced damaging effect was due to the generation of reactive oxygen species (ROS) such as hydroxyl radicals and superoxide radical anions and. Decreased antioxidant markers, together with indomethacin-induced prostaglandins suppression, lead to occlusion of the micro-vessels and subsequent overproduction of ROS metabolites. Especially, indomethacin leads to gastric ulcerations, decreased gluthathione peroxidase enzyme activity and increased lipid peroxidation.

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**Table 2:** Effects of different treatment protocols on the level of various antioxidant markers in the studied rat groups

<table>
<thead>
<tr>
<th>Group</th>
<th>GSH μmol/L</th>
<th>SOD U/ml</th>
<th>CAT μmol/L</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>5.65±0.08</td>
<td>47.76±0.86</td>
<td>232.40±2.88</td>
</tr>
<tr>
<td>IIa</td>
<td>1.02±0.03</td>
<td>22.92±0.64</td>
<td>103.40±2.13</td>
</tr>
<tr>
<td>IIb</td>
<td>3.23±0.02*</td>
<td>39.70±0.68*</td>
<td>186.55±3.47*</td>
</tr>
<tr>
<td>IIc</td>
<td>4.19±0.03**</td>
<td>42.45±0.71**</td>
<td>206.58±4.16**</td>
</tr>
</tbody>
</table>

* Significant difference between MSCs-MVs subgroup (IIb) vs Indomethacin treated subgroup (IIa), \( p < 0.05 \).
# Significant difference between MSCs-MVs subgroup (IIb) vs Alpha-lipoic acid treated subgroup (IIc), \( p < 0.05 \).

**Discussion**

The pathogenesis of Indomethacin-induced gastric ulceration has been attributed to its inhibitory effects on prostaglandin synthesis as well as free radical formation [23]. Indomethacin as one of NSAIDs is frequently used as analgesic. Due to its inherent common side effects and the high cost of gastric ulcer pharmaceuticals, several studies evaluated the effect of natural products of plant origin which proved to be affordable, efficacious and non-toxic. The present study was conducted to evaluate the protective effects of Alpha lipoic acid versus MSC-MVs on indomethacin-induced gastric ulcer in rats.

The histological observations in the ulcerated subgroup IIa revealed areas of loss of superficial epithelium and ulceration. Moreover, there was destruction of the mucosal epithelial cells that showed pyknotic nuclei and highly eosinophilic cytoplasm that extended along the length of fundic glands from the neck to the base. These results agreed with previous studies [24, 25]. Moreover, the antioxidant markers; GSH, SOD and CAT were significantly decreased. These results explain that indomethacin-induced damaging effect was due to the generation of reactive oxygen species (ROS) such as hydroxyl radicals and superoxide radical anions and. Decreased antioxidant markers, together with indomethacin-induced prostaglandins suppression, lead to occlusion of the micro-vessels and subsequent overproduction of ROS metabolites. Especially, indomethacin leads to gastric ulcerations, decreased glutathione peroxidase enzyme activity and increased lipid peroxidation.
Treatment of the ulcerated subgroups with either Alpha lipolic acid or MSCs-MVs showed significant amelioration of the gastric mucosal injury with normal white mucosal rugae and significant decrease in the ulcer index. However, the therapeutic benefits of Alpha lipolic acid significantly exceeded that of MSCs-MVs. This observation could be attributed to the repeated doses of α-Lipoic acid whereas, MSCs-MVs was administered as a single dose.

Our results revealed that the pretreatment with α-Lipoic acid lead to a significant decrease in the ulceration of the stomach suggested by presence of normal appearance of the gastric mucosa with a significantly decreased in ulcer index. Similar results were reported[23,24]. On the other hand, pretreatment with α-Lipoic acid improved the decrease in the antioxidant markers; GSH, SOD and CAT levels which occurred after experimentally induced gastric ulcer. Other studies that used other animal models were showed the antioxidant activities of ALA[25]. Moreover, Kaplan et al[26] stated that α-Lipoic acid protects against gastric mucosal inflammation, oxidations and apoptosis caused by indomethacin by affecting the levels of apoptosis regulator BCL-2-associated protein X (BAX), and by the enhance in the antioxidant system.

As regards effects of injection of microvesicles (MSCs-MVs), our results showed their significant therapeutic effects by amelioration of the gastric mucosal injury induced by indomethacin. Stained sections by HandE of subgroup Ib (MSCs-MVs) were nearly similar in structure as normal gastric mucosa with inflammatory cells infiltration in lamina propria and enlarged gastric pits. Results of transmission electron microscopy (TEM) showed that MSCs-MVs treated group had surface mucous-secreting cells filled with mucin granules of variable size. Some were electron lucent and other electron-dense and had many vacuolations in the cytoplasm. The presence of many dilated intracellular canaliculi in parietal cells suggested their activity. There was also a significant elevation in the antioxidant marker levels (GSH, SOD, CAT) but their levels were not normalized as compared to control group. Immunohistochemical evaluation of VEGF revealed that there was a significant increase in VEGF expression in both subgroups treated with ALA or MSCs-MVs. Our findings coincided with previous studies conducted by Kang et al.[27] and El-Azab et al.[28] who stated that healing of gastric ulcers is accelerated via regulation of VEGF and angiogenesis. Furthermore, MSCs exert significant therapeutic effects in gastric ulcer via secretion of growth factors such as VEGF which induces angiogenesis and preserves the blood supply to gastric mucosa. Stem cells have a significant anti-inflammatory potential by decreasing inflammatory cytokines associated with gastric inflammation as reported by El-Azab et al.[28].

To the best of our knowledge this is the first study conducted to evaluate the effects of microvesicles from MSCs or microvesicles on other pathological types of ulcers such as diabetic foot ulcer.

Li et al.[29] showed that induced pluripotent stem cells derived exosomes (iPS-Exos) exerted a significant therapeutic effect on experimental diabetic ulcer wound healing via promotion of fibroblast migration in vitro and in vivo. Exosomes is a microvesicle that is exocytosed with the phospholipid bilayer and surface antigens. It was reported that exosomes contain miRNA and proteins that could mediate their therapeutic effect by transmitting genetic and non-genetic information communicating cells[30]. Another study conducted on experimental colitis with ulceration showed that Intravenous injection of MSCs-MVs attenuated the severity of colitis as shown by the decrease in the histological colonic damage and the disease activity index. The beneficial therapeutic effects of MSCs-MVs were mediated by modulation of anti-oxidant/ oxidant balance, down regulation of pro-inflammatory cytokines levels, suppression of the apoptosis and inhibition of NF-κBp65 signal transduction pathways[31]. The aforementioned studies stated that exosomes or microvesicles derived from stem cells contain diverse proteins, micro-RNAs, mRNA that could mediate several biological functions and might be the main paracrine mechanism for stem cells-injured cell communication.

In conclusion, Alpha-Lipoic acid (AL) and MSCs-MVs have significant protective effects on gastric mucosa against indomethacin -induced gastric ulcer. Alpha-Lipoic acid showed higher significant antioxidant effect in comparison to the MSCs-MVs, whereas, MSCs-MVs is better than ALA in other parameters so, assessment of the optimal dosage schedule of stem cells derived microvesicles is highly recommended.

CONFLICT OF INTEREST

There are no conflict of interest.

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الملخص العربي

تقييم الاحتمالات العلاجية لحمض ألفا-ليبويك مقابل الحويصلات الصغيرة المشتقة من الخلايا الجذعية الوسيطة ضد قرحة المعدة المستهدفة تجريبياً في الجرذان البيضاء البالغة
(دراسة بالمجهر الضوئي والمجهر الإلكتروني)

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المقدمة: حمض ألفا ليبويك وهو مادة داخلية مضادة للأكسدة. وهو متوفربكميات صغيرة في الغذاء. الحويصلات الصغيرة المشتقة من الخلايا الجذعية الوسيطة لها العديد من الآثار الوقائية على النسيج المصاب نتيجة للأذى التأكسدي أو نتيجة موت الخلايا المبرمج أو الأكسدة.

الأهداف من البحث: أجريت هذه الدراسة لتقييم الاثار الوقائية لحمض ألفا ليبويك مقابل الحويصلات الصغيرة المشتقة من الخلايا الجذعية الوسيطة على قرحة المعدة التي يسببها الإندوميثاسين في الجرذان البيضاء البالغة.

مواد وطرق البحث: تم تقسيم أربعين جردًا من الذكور البالغة إلى: المجموعة الأولى (عشرة جرذان) وتمثل الحيوانات الضابطة، والمجموعة الثانية (ثلاثون جردًا) تضم الجرذان المتقرحة. تلقت المجموعة المصابة بالقرحة جرعة واحدة من الإندوميثاسين (۳ مجم / كجم) وتم تقسيمها إلى: المجموعة الثانية أ (المجموعة المتقرحة)، المجموعة الثانية ب والتي تلقت جرعة واحدة في الوريد من الحويصلات الصغيرة المشتقة من الخلايا الجذعية الوسيطة (۰.۵ مليغ / مل) والتي تلقت ت (ألفا ليبويك ۱۰۰ مليغ / كليل) عن طريق الفم لمدة ۳ أيام قبل استحداث القرحة. تم تقييم الالزيمات المضادة للأكسدة ومؤشر القرحة وأيضا تم فحص أنسجة المعدة بواسطة المجهر الضوئي والمجهر الإلكتروني.

النتائج: تسبب الإندوميثاسين في حدوث أضرار ملحوظة في الغشاء المخاطي في المعدة وكان مؤشر القرحة أعلى بكثير بالمقارنة مع المجموعة الضبطة. كان هناك فقدان للبنية النسيجية الطبيعية، وجود الخلايا الالتهابية. كشف الترتيب الدقيق عن زيادة حبيبات الغشاء المخاطي في خلايا إفراز الغشاء المخاطي. وأظهرت الخلايا الجدارية عن وجود القنائيات داخل الخلايا. أما بالنسبة للجرذان المعالجة بالفليبروك والحوويصلات الصغيرة المشتقة من الخلايا الجذعية الوسيطة كانت هناك زيادة كبيرة في نشاط مضادات الأكسدة كما تم تحسين التركيب الغشائي بشكل كبير مع انخفاض كبير في منطقة قرحة المعدة. وانخفاضات بشكل ملحوظ التتررة والتنخر وجود الخلايا الالتهابية. لم يلاحظ أي ضرر كبير للخلايا البطانية كما انخفض عدد الخلايا الجدارية والخلايا الإيجابية لعامل نمو بطانة الأوعية الدموية بشكل ملحوظ مقارنة مع مجموعة الضبطة.
الاستنتاج: الحويصلات الصغيرة المشتقة من الخلايا الجذعية الوسيطة لها تأثيرات علاجية كبيرة على الغشاء المخاطي في المعدة ضد قرحة المعدة المستحدثة باستخدام الإندوميثاسين عن التأثير الوقائي لحمض ألفا ليبويك.